

CENTRAL UNIVERSITY OF PUNJAB



M.Sc. in Molecular Medicine

Batch – 2024-2026

**Department of Human Genetics and
Molecular Medicine**

Syllabus

MSc in Molecular Medicine (Batch: 2024-2026)

Graduate Attributes

Context of Society

The students of this course will understand the significant role of cell and molecular biology, Pathophysiology of common diseases and therapeutic strategies in the broader societal context. There will be an understanding of the basics of professional ethics, research ethics, biosafety issues, and the principles of professional practice.

Enterprising and Knowledgeable

The course content will develop skills in regenerative medicine, molecular biology, and bioinformatics and cutting-edge molecular techniques through the subject content across a broad range of modules among the students. The development of skills in molecular medicine will enhance employability in the field biomedical sciences and clinical practices. The emphasis is on student-centric learning where they solve the patterns of inheritance by drawing pedigrees and discuss the current therapeutic interventions to treat specific genetic disorders.

Digital and research methodology-based skills

The students will be able to study and learn the effective use of digital tools to support academic writing, reference management and independent study using digital resources and learning materials. The understanding of the principles of experimental design and methods will help the students to explore new scientific approaches in translational research.

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Course Structure of the Programme

Total Credit: 85

	Core Subjects	Elective Courses			Foundation Courses		Total Credit
		DE	ID	SB	CF	EF/VB	
Sem-I	03 (9 Cr)	01 (3 Cr)	--	03 (4 Cr)	01 (3 Cr) 01 (3 Cr)	--	22
Sem-II	04 (12 Cr)	01 (3 Cr)	01 (2 Cr)	01 (3 Cr)	--	--	20
SEM-III	03 (9 Cr)	01 (3 Cr)	--	01 (3 Cr) 01 (4 Cr Dissertation)	02 (2 Cr)	01 (2 Cr)	23
SEM-IV	--	--	--	01 (20 Cr Dissertation)	--	--	20
Credit Score	30	9	02	34	8	02	85

DE: Discipline Elective

ID: Interdisciplinary

SB: Skill based (Practicals); Dissertation

CF: Compulsory foundation

EF: Elective Foundation

VB: Value Based

Note:

- i. Each semester will include a 01 non-credit course (2 hour) on individualized education plan/tutorials.
- ii. **Post-graduate Diploma in Molecular Medicine** will be awarded to students who **Exit** after successful completion of first two semesters along with skill development course (6 Cr.) at the University level.
- iii. **Re-entry** will only be allowed to those who successfully completed the **Post-Graduate Diploma in Molecular Medicine**.

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Course Structure of the Programme Semester-I

Course Code	Course Title	Course Type	Hours			Credit
			L	T	P	
MME.506	Cell Biology	Core	3	0	0	3
HGE.507	Concepts of Genetics	Core	3	0	0	3
MME.508	Biomolecules and Metabolism	Core	3	0	0	3
HGE.509	Biostatistics and Research Methodology	Compulsory Foundation	3	0	0	3
MME.510	Molecular Biology	Compulsory Foundation	3	0	0	3
HGE.511	Concepts of Genetics (Practical)	Skill Based	0	0	2	1
MME.512	Biomolecules and Metabolism (Practical)	Skill Based	0	0	4	2
HGE.513	Biostatistics and Research Methodology (Practical)	Skill Based	0	0	2	1
Discipline Elective Course-I (Any one of the following)						
HGE.515	Public Health Research and Genetic Epidemiology	DE	3	0	0	3
MME.515	Molecular and Cellular Oncology	DE	3	0	0	3
MIC.525	Microbial Pathogenicity	DE	3	0	0	3
ZOL.528	Basics in Neuroscience	DE	3	0	0	3
Total Credits						22

- *Additional: 01 non-credit course (2 hr) on individualized education plan/tutorials.*

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Semester-II

Course Code	Course Title	Course Type	Hours			Credit
			L	T	P	
MME.521	Stem Cell and Regenerative Medicine	Core	3	0	0	3
MME.522	Essentials of Immunology	Core	3	0	0	3
MME.523	Techniques in Molecular Medicine	Core	3	0	0	3
HGE.524	Human Anatomy and Physiology	Core	3	0	0	3
MME.525	Practical Course in Molecular Medicine	Skill Based	0	0	6	3
Interdisciplinary Course-I (For other Departments)						
HGE. 529	Introduction to Intellectual Property Rights	IDC	2	0	0	2
xxx	MOOC course	IDC	2	0	0	2
XXX	Choose from IDC courses offered by other Departments	IDC	2	0	0	2
Discipline Elective Course-II (select anyone)						
MME.527	Molecular Endocrinology and Signal Transduction	DE	3	0	0	3
HGE.527	Human Embryology and Developmental Genetics	DE	3	0	0	3
BCH.528	Secondary Metabolites and Xenobiotic Metabolism	DE	3	0	0	3
BIM.521	Big Data Analysis in Healthcare	DE	3	0	0	3

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	Total Credits 20
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- *Additional: 01 non-credit course (2 hr) on individualized education plan/tutorials.*
- *Additional options for MOOC course may be given to students based on the availability of actively running/newly introduced MOOC courses in SWAYAM platform at the time of course selection. Approval for the same may be granted by the HoD”*

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Semester-III

Course Code	Course Title	Course Type	Hours			Credit
			L	T	P	
MME.551	Molecular Basis of Human Diseases	Core	3	0	0	3
MME.552	Genetic Engineering and Recombinant Therapeutics	Core	3	0	0	3
MME.553	Trends in Molecular Medicine	Core	3	0	0	3
HGE.554	Tools in Bioinformatics (Practical)	Skill Based	0	0	6	3
Discipline Elective Course-III (select anyone)						
MME.555	Evolution and Developmental Biology	DE	3	0	0	3
HGE.555	Biosafety, Bioethics and IPR	DE	3	0	0	3
MIC.559	Microbial Biotechnology	DE	3	0	0	3
BCH.557	Clinical Diagnostics	DE	3	0	0	3
Value Added Course Courses						
HGE.556	Principles of Ecological Sciences	Value based	2	0	0	2
XXX	Value Added Course (From other departments)	VAC	2	0	0	2
Compulsory Foundation Course						
HGE.558	Innovation and Entrepreneurship	Compulsory Foundation	2	0	0	2
MME.600	Dissertation Part-I	Skill Based	0	0	8	4
Total						23

- Additional: 01 non-credit course (2 hr) on individualized education plan/tutorials.

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Semester-IV

Course Code	Course Title	Course Type	Hours			Credit
			L	T	P	
MME.601	Dissertation Part-II	Skill Based	0	0	40	20
Total			0	0	40	20

MOOC Options: 40% credits can be obtained through MOOCs depending on availability of course with matching content.

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Semester I

Course Code: MME.506
Course Title: Cell Biology
Total Hours: 45

L	T	P	C
3	0	0	3

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Understanding structures and functions of various cellular organelles.

CLO2: Conceptualization of basic cellular mechanisms.

CLO3: Conceptualize the mechanisms of inter- as well as intra-cellular communications.

CLO4: Understanding the cell cycle regulation and its importance in disease biology

Unit 1 Introduction to the cell: Models of membrane structure, Membrane proteins, Membrane carbohydrates, Membrane transport of small molecules, Membrane transport of macromolecules and particles. Structural organization and function of intracellular organelles: The lysosomes, Ribosomes, The peroxisomes, The Golgi apparatus, The endoplasmic reticulum, Mitochondria.	12 Hours	CLO1
Unit 2 Protein secretion and sorting: Protein secretion, synthesis and targeting to mitochondria, chloroplast, peroxisomal proteins, translational modification in the ER. Intracellular traffic, vesicular traffic in the secretory pathway, protein sorting in the Golgi bodies, traffic in the endocytic pathway, exocytosis.	10 Hours	CLO2
Unit 3 The cytoskeleton: The nature of cytoskeleton, Intermediate filaments, Microtubules, Actin filaments, Cilia and centrioles, Organization of the cytoskeleton. Cell communication and cell signaling: Cell adhesions, Cell junctions and the extra cellular matrix, Cell-cell adhesion and communication, Cell matrix adhesion, Collagen the fibrous protein of the matrix, Non-collagen component of the extra cellular matrix.	14 Hours	CLO3
Unit 4 Cell growth and division: Overview of the cell cycle and its control, molecular mechanisms for regulating mitotic and meiotic events, Amitosis, Cell cycle control, Checkpoints in cell cycle regulation.	9 Hours	CLO4

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Transactional Modes: Lecture; Demonstration; Tutorial; Lecture cum demonstration; Problem solving; Self-learning.

Internal assessment shall be through any of the following: Surprise Tests, one sentence summary, case analysis, simulated problem solving, open book techniques, classroom assignments, homework assignments, term paper, presentations, and discussions.

Suggested Reading:

1. Alberts, B., Heald, R., Johnson, A., Morgan, D., Raff, M. Roberts, K., and Walter, P. (2022). Molecular Biology of the cell. Garland publishers, Oxford.
2. Celis, J.E. (2006). Cell biology: A laboratory handbook, Vol 1, 2, 3. Academic Press, UK.
3. Gupta, P.K. (2018). Cytology, Genetics and Evolution. Rastogi publications, Meerut, India.
4. Karp, G. (2022). Cell and Molecular Biology: Concepts and Experiments. John Wiley & Sons. Inc. New Delhi, India.

Course Code: HGE.507
Course Title: Concepts of Genetics
Total Hours: 45

L	T	P	C
3	0	0	3

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Evaluate the Mendelian and Non-Mendelian inheritance patterns. Gather knowledge about gene expression regulation and sex determination,

CLO2: Evaluate different chromosomal aberrations and ploidies

CLO3: Know about genetic system of microbes

CLO4: Know the details of extra chromosomal inheritance patterns

Unit I	11 Hours	CLO1
Transmission Genetics: Mendel's laws of inheritance and its applications; concept of segregation, independent assortment and dominance; pedigree analysis; epistasis; crossing over and recombination; gene linkage and genetic mapping Sex determination: Sex determination in Human and Drosophila; X-chromosome inactivation; dosage compensation.		

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Unit II 11 Hours Chromosomal Aberrations and Gene mutations: Numerical aberrations: aneuploidy and polyploidy; structural chromosomal aberrations: deletion, duplication, inversion and translocation, types of mutations: point mutation, transitions and transversions, physical and chemical mutagens, Benzer experiment to elucidate fine structure of gene; Chromosomal changes and evolution of bread wheat.	11	CLO2
Unit III Hours Extra-chromosomal inheritance: Chloroplast: variegation in Four O' Clock plants; cytohets of <i>Chlamydomonas</i> ; mitochondrial inheritance: poky in neurospora, petites in yeast; infectious heredity: Kappa in <i>Paramecium</i> , infective particles in <i>Drosophila</i> .	11	CLO3
Unit IV Hours Population Genetics Population Dynamics, conditions and deviations of the Hardy-Weinberg law; selection coefficient and fitness; heterozygous advantages; inbreeding and its consequences; mutation pressure and genetic load; dynamics of migration and genetic drifts.	11	CLO4

Transactional Modes: Lecture; Demonstration; Tutorial; Lecture cum demonstration; Problem solving; Self-learning; co-learning techniques.

Internal assessment shall be through any of the following: Report on novel chromosome aberrations in human, Discussion on transposable elements in human diseases, Surprise Tests, one sentence summary, case analysis, simulated problem solving, open book techniques, classroom assignments, homework assignments, term paper, Karyotype analysis presentations and discussions.

Suggested Readings:

1. Klug WS and Cummings MR. Concepts of Genetics. Prentice-Hall.2014
2. Anthony, J.F. Miller, J.A. Suzuki, D.T., Richard, R.C., Gilbert, W.M. (1998). An introduction to Genetic Analysis. W.H. Freeman publication, USA.
3. Pierce BA. Genetics: A Conceptual approach. Freeman Publishers.
4. Hartle DL and Jones EW. Genetics: Analysis of Genes and Genomes. Jones & Bartett.
5. Atherly, A.G., Girton, J.R., Mcdonald, J.F. (1999). The Science of Genetics. Saundern College publication.
6. Snusted, D.P., Simmons, M. J. (2010). Principles of Genetics. John Wiley & Sons, New
7. Griffith AF et al. An Introduction to Genetic Analysis. John Wiley & Sons.

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Course Code: MME.508
Course Title: Biomolecules and Metabolism
Total Hours: 45

L	T	P	C
3	0	0	3

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Evaluate the basic concepts of Enzymes, Enzyme Kinetics, and its role in metabolic processes. Conceptualize the basic features of enzyme catalysis and regulation.

CLO2: Study the basic structural features of carbohydrates and its metabolism.

CLO3: Conceptualization of pathways in lipid metabolism.

CLO4: Study the structure of nucleic acids, amino acids and proteins and their metabolism.

<p>Unit-1 Enzymes: Importance & Classifications; Properties of enzymes; Isozymes; Enzyme nomenclature; Factors affecting enzyme action, Enzyme regulation; Mechanism of enzyme action; Enzyme kinetics & enzyme inhibitors; ES complex formation; Michaelis-Menten equation; Line-weaver Burk plot; Km value and its significance; Turnover number; Enzyme inhibitors, Allosteric enzymes. Ribozymes and Abzymes.</p>	12 Hours	CLO1
<p>Unit-2 Hours Carbohydrates: Structure and functions of monosaccharides, disaccharides, and polysaccharides; Epimers; Anomers; mutarotation. Reactions of carbohydrates. Carbohydrate's metabolism - General scheme of metabolism. Glycolysis, TCA cycle, Gluconeogenesis, Glycogenolysis, Pentose phosphate pathway, and their regulation; Oxidative phosphorylation</p>	11	CLO2
<p>Unit-3 Lipids: Classification - simple, compound and derived lipids with examples and their role in the human body. Lipid metabolism: Beta oxidation - Pathway and regulation. Role of acyl carnitine in fatty acyl transport. Biosynthesis of cholesterol and regulation. Formation of Ketone bodies.</p>	11 Hours	CLO3

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Unit-4 Amino acids: Metabolism of Amino Acids. Proteins: Classification and biological importance of protein in human body; Secondary, Tertiary and Quaternary structure, Ramachandran plot. Oxygen binding proteins – Hemoglobin and myoglobin. Hill equation, Bohr’s Effect. Nucleic Acids: Metabolism of purines and pyrimidines- Salvage and de novo pathways.	11 Hours	CLO4
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Transactional Modes: Lecture; Demonstration; Tutorial; Lecture cum demonstration; Problem solving; Self-learning.

Internal assessment shall be through any of the following: Surprise Tests, Group discussion, one sentence summary, case analysis, simulated problem solving, open book techniques, classroom assignments, homework assignments, term paper, presentations, and discussions.

Suggested Readings:

1. Berg, J. M., Tymoczko, J. L., & Stryer, L. (2012). Biochemistry. W.H. Freeman & Company. USA.
2. Brown, T. A. (2016). Gene cloning and DNA analysis: an introduction. John Wiley & Sons.
3. Price, N. C., Price, N. C., & Stevens, L. (2003). Fundamentals of Enzymology: The Cell and Molecular Biology of Catalytic Proteins. Oxford University Press.
4. Mathews, C.K., Van Holde, K.E. & Ahern, K.G. (2000). Biochemistry. Oxford University Press Inc. New York.
5. Nelson, D. & Cox, M.M. (2008). Lehninger Principles of Biochemistry. BI publications Pvt. Ltd. Chennai, India.
6. Palmer, T., & Bonner, P. L. (2001). Enzymes: Biochemistry. Biotechnology, Clinical Chemistry. Horwood Publishing Chichester.
7. Deininger, P. (1990). Methods in enzymology, vol. 185, gene expression technology: Edited by David V. Goeddel. Academic Press, San Diego, CA.
8. Raven, P.H., Johnson, G.B. & Mason, K.A. (2007). Biology. Mcgraw-Hill. USA.
9. Shukla, A. N. (2009). Elements of enzymology. Discovery Publishing House.
10. Voet, D. & Voet, J.G. (2008). Principles of Biochemistry. CBS Publishers & Distributors. New Delhi, India.
11. R Swaminathan. (2011). Handbook of clinical biochemistry. 2 edition, World Scientific Publishing Company, New Jersey, USA

Course Code: HGE.509

Course Title: Biostatistics and Research Methodology

Total Hours: 45

L	T	P	C
3	0	0	3

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Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Analyze and evaluate wide variety of statistical data

CLO2: Compose statistical data and summary statistics in graphical and tabular forms. Perform biological sampling and statistical analysis.

CLO3: Apply suitable statistical tools to analyze data

CLO4: Write and communicate scientific reports, projects, and publications.

UNIT I Overview of Biostatistics: Basic concepts of statistical data and different types of tables; graphical representation of experimental data for publication; frequency distribution; measurement of central tendency and variation.	11 Hours	CLO1
UNIT II Experimental design and analysis: Basics of sampling in biological studies; different types of sampling techniques; various steps in sampling; concept of data distribution in sampling; graphical representation of data; level of significance; hypothesis testing, Errors.	11 Hours	CLO2
UNIT III Inferential Statistics: Chi-Square test: hypothesis testing, contingency, homogeneity; student's t-test: paired and unpaired, one tailed and two tailed; one-way and two-way analysis of variance (ANOVA); correlation and regression.	11 Hours	CLO3
UNIT IV Study design & Technical writing: Best practices in research and technicality of research design; interpretation and report writing, e-Library; web-based literature search engines; evaluation-based development of scientific writing skill: synopsis, research paper, poster preparation and paper presentation and dissertation.	12 Hours	CLO4

Transactional Modes: Lecture; Tutorial; Problem solving; Self-learning.

Internal assessment shall be through any of the following: Surprise Tests, student generated questions, case analysis, simulated problem solving, open book techniques, classroom assignments, homework assignments, term paper.

Suggested Readings:

1. Rao, S.P.S.S. Richard, J. (2016). Introduction to biostatistics and research methods. 5th Ed. Phi Learning Pvt. Ltd. New Delhi.
2. Hoffman, J. (2019). Biostatistics for medical and biomedical practitioners. 2nd Ed. Academic Press, London.
3. Banerjee P.K (2014). Introduction to Biostatistics. S.Chand, New Delhi.
4. Antonisamy, B. Christopher, S. Samuel, P.P. (2011). Biostatistics: Principles and Practice. Tata McGraw Hill. New Delhi.

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5. Daniel W.W (2013). Biostatistics: Basic Concepts and methodology for the health sciences. 10th Ed. John Wiley and Sons Inc, New Delhi.
6. Norman, G. and Streiner, D. (2008). Biostatistics: The Bare Essentials. (With SPSS), 3rd Edition, Decker Inc. USA.
7. Sokal, R.R. and Rohlf, F.J. (1994). Biometry: The Principles and Practices of Statistics in Biological Research. W.H. Freeman publishers, USA.

Course Code: MME.510
Course Title: Molecular Biology
Total Hours: 45

L	T	P	C
3	0	0	3

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Conceptualization of molecular mechanisms involved in cellular functioning.

CLO2: Understanding the molecular processes of DNA replication

CLO3: Study the process of transcription in prokaryotes and eukaryotes.

CLO4: Understand the concept of Genetic code, process of translation and post translational modifications

Unit 1 Hours Nucleic acids, Genes and Genome organization: Chemical structure of DNA and base composition, Watson-Crick model, mitochondrial DNA, Chromosome Structure, Chromatin and the Nucleosome, Chromatin structure: euchromatin, heterochromatin, Constitutive and facultative heterochromatin, Regulation of chromatin structure and nucleosome assembly, typical structure of a eukaryotic genes including various regulatory elements.	12	CLO1
Unit 2 DNA replication and repair: Mechanisms of DNA replication in eukaryotes, Enzymes and accessory proteins involved in DNA replication, Replication errors and proofreading, telomeres, DNA damage and repair mechanisms.	12 Hours	CLO2
Unit 3 Transcription and mRNA processing: Different forms of RNA: mRNA, tRNA, rRNA and other Types of RNA Eukaryotic transcription: Initiation, Elongation & Termination, general and specific transcription factors, Regulatory elements and mechanisms of transcription regulation, RNA processing and editing, post transcriptional gene regulation.	11 Hours	CLO3
Unit 4 Translation: Genetic code, eukaryotic translation, the translation machinery, mechanisms of chain initiation, elongation and termination, regulation of translation, co- and post- translational modifications of proteins.	10 Hours	CLO4

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Transactional Modes: Lecture; Demonstration; Tutorial; Lecture cum demonstration; Problem solving; Self-learning.

Internal assessment shall be through any of the following: Surprise Tests, one sentence summary, case analysis, simulated problem solving, open book techniques, classroom assignments, homework assignments, term paper, presentations and discussions.

Suggested Reading:

1. Sambrook, J., Fritsch, E. F., & Maniatis, T. (2015). Molecular cloning: a laboratory manual. Cold Spring Harbor Laboratory Press New York.
2. Berk, A. Chris, A.K. & Krieger, M. (2021). Molecular Cell Biology. W.H. Freeman, USA.
3. Robertis, (2017). Cell and Molecular Biology. Lippincott Williams & Wilkins.
4. Karp, G. (2019). Cell and molecular biology: concepts and experiments. John Wiley & Sons.
5. Krebs, J. E., Goldstein, E. S., & Kilpatrick, S. T. (2017). Lewin's Genes XII. Jones & Bartlett Learning.
6. Watson, J. D., Baker, T. A., Bell, S. P., Gann, A., & Levine, M. (2007). Molecular Biology of the Gene Benjamin Cummings.
7. Alberts, B., Johnson, A., Lewis, J., Raff, M., Roberts, K., & Walter, P. (2022). Molecular biology of the cell. Garland Science. New York, 1392.
8. Fasman, G.D. (1989). Practical Handbook of Biochemistry and Molecular Biology. CRC Press, Taylor and Francis Group, UK.
9. Cooper GM. The Cell: A Molecular Approach. 2nd edition. Sunderland (MA): Sinauer Associates; 2019.

Course Code: HGE.511

Course Title: Concepts of Genetics (Practical)

Total Hours: 30

L	T	P	C
0	0	2	1

Course Learning Outcomes:

On successful completion of the course the student will be able to:

1. Perform any experiments on Mendelian genetics and pedigree analysis
2. Perform linkage based genetic analysis
3. Design genetic experiments using common model organisms
4. Isolate genomic DNA for genetic analyses

List of Practical work:

1	Problems on Monohybrid and dihybrid ratios, Multiple alleles, Epistasis	CLO1
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2	Inheritance patterns in Human– Numerical on Pedigree analysis- Autosomal patterns, X–linked patterns, Y–linked patterns	
3	Segregation analysis in Drosophila (Monohybrid, Dihybrid)	
4	Analysis on Linkage	CLO2
5	Linkage mapping	
6	Identification of inactivated X chromosome as Barr body	CLO3
7	Studies of a Model organism: <i>E. coli</i> , <i>C.elegans</i> , <i>D. melanogaster</i> and <i>D. rerio</i>	
8	Isolation of genomic DNA and quality control	CLO4

Transactional Modes: Hands-on practical's; Demonstration; Virtual classrooms; Tutorial; Self-directed learning.

Evaluation criteria for practical courses:

- Continuous Assessment = **60 Marks**

Based on performance of the students and completion of the record book (60 divided by the number of practicals)

- Final Examination = **40 Marks**

Subjective question = **10 Marks**

Performing experiment = 20 Marks

Viva voce = 10 Marks

Course Code: MME.512

Course Title: Biomolecules and Metabolism (Practical)

Total Hours: 60

L	T	P	C
0	0	4	2

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Prepare buffers and solutions with varied concentration.

CLO2: Isolation of DNA from the blood samples

CLO3: Quantitative estimation of biomolecules and their role in health and disease

CLO4: Evaluate the effect of temperature, pH and substrate concentration on enzyme activity

List of Practicals

1. Preparation of Buffers	CLO1
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2. Extraction of DNA, and purity check by electrophoresis.	CLO2
3. Quantitative estimation of Glucose. 4. Quantitative estimation of Cholesterol 5. Quantitative estimation of Proteins 6. Quantitative estimation of Nucleic Acids	CLO3
7. Assay of enzyme activity in saliva. 8. Effect of temperature on enzyme activity. 9. Effect of pH on enzyme activity.	CLO4

Suggested Readings:

1. Rajendran, S., Dhiman, P. (2019) Biochemistry Practical Manual. Elsevier India.
2. Plummer, D. (2004) An Introduction to Practical Biochemistry. Tata McGraw Hill Publishers Co. Ltd., India
3. Gupta, P.P., Gupta, N. (2017) Essentials of Practical Biochemistry. Jaypee Brothers Medical Publishers Pvt. Ltd. India.
4. Hofmann, A., Clokie, S. (2018). Wilson and Walker's Principles and Techniques of Biochemistry and Molecular Biology. Cambridge University Press, USA
5. Murphy M.J., Srivastava R., Deans, K. (2018) Clinical Biochemistry, 6th Edition. Elsevier's USA.
6. Swaminathan, R (2011) Handbook of Clinical Biochemistry, 2nd Edition, Publisher: World Scientific

Transactional Modes: Laboratory based practical; Problem solving; Self-learning.

Internal assessment shall be through any of the following: Case analysis, Lab Performance, Open book techniques, Instruments Demonstration, and Group discussions.

Evaluation criteria:

- A. Continuous Assessment = **60 Marks** (Based on performance and good lab practices of the students and completion of the record book (60 divided by the number of practicals))
- B. Final Examination = **40 Marks**
- i. Subjective question = 10 Marks
 - ii. Performing experiment = 20 Marks
 - iii. Viva voce = 10 Marks

Course Code: HGE.513

Course Title: Biostatistics and Research Methodology
(Practical)

L	T	P	C
0	0	2	1

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Total Hours: 30

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Identify statistical data type and plot graphs using conventional tools. Perform basic statistics to check the data quality

CLO2: Test correlation and regression using two or more variables

CLO3: Perform standard parametric and non-parametric statistics on simple data

CLO4: Write and communicate scientific literatures.

List of Practical work:

1	Plotting different types of graphs and statistical tables using MS excel, GraphPad and/or relevant tools	CLO1
2	Plotting normal distribution from data given	
3	Problems on: central tendency, measurement of variance (standard deviation, standard error etc.)	
4	Problems of correlation	CLO2
5	Problems of regression	
6	Chi-square test	CLO3
7	Student's t-test	
8	Analysis of variance (ANOVA)	
9	Different techniques of sampling	
10	Scientific writing skill development	CLO4
11	Practice writing research reports, synopsis, poster etc.	

Transactional Modes: Laboratory based practical's; demonstration, Problem solving; Self-learning.

Evaluation criteria for practical courses:

- Continuous Assessment = **60 Marks**

Based on performance and good lab practices of the students and completion of the record book (60 divided by the number of practical's)

- Final Examination = **40 Marks**

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- i. Subjective question = 10 Marks
- ii. Performing experiment = 20 Marks
- iii. Viva voce = 10 Marks

Discipline Elective Courses (Select one)

Course Code: HGE.515
Course Title: Public Health Research and Genetic Epidemiology
Total Hours: 45

L	T	P	C
3	0	0	3

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Develop applicable knowledge on global and local public health issues.

CLO2: Perform public health data management and result interpretation.

CLO3: Perform different epidemiological studies to identify the cause-effect relationship in variety of human traits/diseases

CLO4: Design genetic studies and perform association and linkage analysis on any relevant data.

Unit I Hours Principles and Practices of Public Health: Scope and concerns of public health; Determinants and measurement of health and disease; Environment and health; Health equity.	11	CLO1
Unit II Hours Public health management: Occupational health; Exposure and risk management; Health informatics; Concepts of Genomic surveillance; Clinical trials.	11	CLO2
Unit III Fundamentals of epidemiological studies: Experimental and observational studies; Cohort study; Cross-sectional studies; Methods of sampling; Methods of association studies; Genome-wide association studies (GWAS). Systematic review and meta-analysis.	12 Hours	CLO3
Unit IV Application of genetic variations: Basics of genetic variations; genetic markers – SNP, CNV, Ins/dels, VNTR, STR, microsatellite; concepts of tag markers and haplotypes; linkage disequilibrium; quantitative genetic analysis; QTL and eQTL.	11 Hours	CLO4

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Transactional Modes: Lecture; Demonstration; Tutorial; Innovative ideation; Case study; Virtual classrooms; seminar presentation; Problem solving; Co-learning techniques; Self-learning.

Internal assessment shall be through any of the following:

Surprise Tests, one sentence summary, case analysis, simulated problem solving, open book techniques, classroom assignments, homework assignments, term paper, Seminars, term papers, discussions and presentations.

Suggested Readings:

1. Detela, R. Gulliford, M. Karim, Q.A. Tan, C.C. (2021). Global Public Health. Oxford University Press. 7th Ed.
2. Park, K. (2021). Preventive and Social Medicine. Bhanot Publishers. New Delhi.
3. U.S Department of Health and Human Services. (2012). Principles of Epidemiology in Public Health Practice. Updated Third Ed.
4. World Health Organization. (2001). Health Research Methodology: A guide for training in research methods. 2nd Ed.
5. Celentano, D.D. and Szklo, M. (2019) Gordis Epidemiology. Elsevier. 6th Ed.
6. Palmer LJ, Burton PR & Smith GD (2011). An introduction to genetic epidemiology (Policy Press, University of Bristol)
7. Dawn TM (2011). Genetic Epidemiology (Springer)
8. Austin M (2013). Genetic Epidemiology: Methods and Applications, 1st Edition (CABI Publishing).

Course Code: MME.515

Course Title: Molecular and Cellular Oncology

Total Hours: 45

L	T	P	C
3	0	0	3

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Understand fundamentals of cancer.

CLO2: Gain knowledge about genetics and signal transduction involved in tumorigenesis.

CLO3: Learn about various tools used for diagnostic purposes.

CLO4: Understand basics principles of anticancer therapeutics as well as about recent developments of the field.

Unit: 1 Hours Fundamentals and Genetics of Cancer: Hallmarks of cancer, cancer classification, Mutagens, carcinogens and gene mutations, Chromosomal aberrations, tumor viruses and discovery of oncogenes, tumor suppressors and oncogenes, familial cancer syndromes, telomere regulation in cancer.	13	CLO1
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Unit: 2 Hours Signal Transduction in Cancer Progression: Deregulation of Cell cycle in cancer. Cell signaling in cancer; cancer metabolism; hypoxia and metastasis, angiogenesis, tumor microenvironment. DNA damage and repair defects and their relation to cancer, cancer stem cells	11	CLO2
Unit: 3 Hours Cancer Detection: General and organ specific symptoms associated with cancer, techniques for cancer detection, biomarkers for cancer detection of various stages of cancer, In-vitro assays to detect angiogenesis, metastasis, cell proliferation, mice models to study cancer (transgenic, knock-out, knock-in, xenografts and patient derived xenografts), genomic and proteomic approaches to develop better cancer markers.	11	CLO3
Unit: 4 Hours Cancer Therapies and Recent Advances in Cancer Research: Traditional Chemotherapies, radiotherapy, Onco-surgery, Bone marrow transplantation, stem cell therapies, Immunotherapy, combinational therapies, natural products as therapeutics, cancer vaccines, gene therapies, targeted anticancer therapies, CAR T and Indian traditional medicine for cancer therapies.	10	CLO4

Transactional Modes: Lecture; Demonstration; Tutorial; Lecture cum demonstration; Problem solving; Self-learning.

Internal assessment shall be through any of the following: Surprise Tests, case analysis, simulated problem solving, classroom assignments, term paper, presentations and discussions.

Suggested Reading:

1. DeVita, V. T., Rosenberg, S. A., & Lawrence, T. S. (2018). DeVita, Hellman, and Rosenberg's cancer. Lippincott Williams & Wilkins.
2. Enders, G. H. (2010). Cell cycle deregulation in cancer. Humana Press, Springer science, New York.
3. Gusev, Y. (2019). Micro RNA Profiling in Cancer. Pan Stanford publishing pvt.Ltd., Singapore.
4. Hiem, S., & Mitelman, F. (2019). Cancer Cytogenetics. IIIrd edition. Willey-Blackwell publishing, New Jersey.
5. Jocelyn, E. K., Elliot, S. G., Stephen, T. K. (2018). Lewin's Gene X. Jones & Barlett.
6. Wang, E. (2018). Cancer systems biology. CRC press, Taylor & Francis group, New York.
7. Jacques Robert (2015) Textbook of cell signalling in cancer, Springer science, New York.

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8. Weinberg, Robert A. (2023), 3rd edition, The Biology of Cancer. New York: Garland Science

Related Weblink

<http://www.insidecancer.org/>

<http://www.who.int/cancer/en/>

<http://www.cancer.gov/>

http://www.icmr.nic.in/ncrp/cancer_reg.htm

Semester-II

Course Code: MME.521

Course Title: Stem Cell and Regenerative Medicine

Total Hours: 45

L	T	P	C
3	0	0	3

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Understand basic stem cell biology.

CLO2: Understand basics of cell culture

CLO3: Gain conceptual knowledge about requirements for tissue engineering.

CLO4: Know regenerative medicine and its potential applications.

Unit: 1 Hours Stem Cells: Stem cells and their properties, classification of stem cells, in-vitro culture techniques, isolation, identification and characterization of stem cells, stem cells in various organs and in disease conditions; Cutting-Edge Research in Stem Cell Biology	11	CLO1
Unit: 2 Hours Cell Culturing: Basics of cell culture and media, Culturing primary cells and cell lines, suspension and adherent cultures, cell growth, growth inhibition and apoptotic studies, Embryo culture, transplantation and teratogens, teratomas, stem cell culture, organ culture.	12	CLO2

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Unit: 3 Hours Tissue Engineering: Principles of tissue culture, tissue and organ culture, extracellular matrices, bioreactors, ethical issues related to stem cell therapies, stem cell banks, bone marrow transplantation; Tissue regeneration and repair-3D bioprinting.	11	CLO3
Unit: 4 Hours Regenerative Medicine: Modes of tissue and organ delivery, tissue Engineering and transplantation techniques, immuno isolation techniques, regeneration of bone and cartilage, Islet cell transplantation and bio-artificial pancreas, lung regeneration	11	CLO4
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Transactional Modes: Lecture; Demonstration; Tutorial; Lecture cum demonstration; Problem solving; Self-learning.

Internal assessment shall be through any of the following: Group discussions, Surprise Tests, case analysis, simulated problem solving, classroom assignments, term paper, presentations and discussions.

Suggested Reading:

1. Lanza, R., Gearhart, J. (2016). Essential of Stem Cell Biology. Elsevier Academic Press.
2. Lanza, R., Klimanskaya, I. (2018). Essential Stem Cells Methods. Academic Press.
3. Mao, J. J. & (2017). Translational approaches in tissue engineering and regenerative medicine. Artech House.
4. Lanza, R. (2017). Principles of Tissue Engineering, 3rd Edition. Academic Press
5. Stein, G. S., Borowski, M., Luong, M. X., Shi, M. J., Smith, K. P., & Vazquez, P. (Eds.). (2011). Human stem cell technology and biology: A research guide and laboratory manual. John Wiley & Sons.
6. Lanza, R., Blau, H., Gearhart, J., Hogan, B., Melton, D., Moore, M., ... & Weissman, I. (Eds.). (2014). Handbook of Stem Cells, Two-Volume Set: Volume 1-Embryonic Stem Cells; Volume 2-Adult & Fetal Stem Cells. Elsevier.

Course Code: MME.522
Course Title: Essentials of Immunology

L	T	P	C
3	0	0	3

Syllabus

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Total Hours: 45

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Evaluate basic concepts of immune system.

CLO2: Gain knowledge about various key processes related to development of immune system.

CLO3: Understand the concept of immune-based diseases as either a deficiency of components or excess activity as hypersensitivity.

CLO4: Apply the knowledge how immune system is involved in diseases caused by internal or external factors.

Unit: 1 Hours Immune System: The cells and organs of immune system, humoral immunity-immunoglobulin, basic structure, classes and subclasses, structural and functional relationships, nature of antigen, antigen-antibody reaction, antibody diversity, class switching, B and T cell development.	12	CLO1
Unit: 2 Hours Immune Effectors: Complement system, their structure, functions and mechanisms of activation by classical, alternative and lectin pathway. Th1 and Th2 response, various effector cells of the immune system: DC, NK, Monocytes etc.	11	CLO2
Unit: 3 Hours Mechanisms of Immune System Diversity: Structure and functions of Major Histocompatibility Complex (MHC) and Human Leukocyte Antigen (HLA) system, polymorphism, distribution, variation and their functions.	12	CLO3
Unit: 4 Hours Immune System in Health and Diseases: Inflammation, hypersensitivity and autoimmunity, AIDS and immunodeficiency, Transplantation immunology , vaccine development.	10	CLO4

Transactional Modes: Lecture; Demonstration; Tutorial; Lecture cum demonstration; Problem solving; Self-learning.

Internal assessment shall be through any of the following: Surprise Tests, one sentence summary, case analysis, simulated problem solving, open book techniques,

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classroom assignments, homework assignments, term paper, presentations and discussions.

Suggested Reading:

1. Kindt, T.J., Osborne, B.A. and Goldsby, R.A. (2018). Kuby Immunology. W.H. Freeman, USA.
2. Abbas. (2018). Cellular and Molecular Immunology. CBS Publishers & Distributors, India.
3. Charles, A. and Janeway, J.R. (2001). Immunobiology: The immune system in health and disease Blackwell Publishing, USA.
4. Delves, P.J., Roitt, I.M. and Seamus, J.M. (2016). Roitt's Essential Immunology (Series–Essentials). Blackwell Publishers, USA.
5. Elgert, K.D. (2015). Immunology: Understanding the immune system. Wiley-Blackwell, USA.
6. Tizard. (2018). Immunology: An Introduction. Cengage Learning, Thompson, USA.
7. Owen, Judith A; Punt, Jenni, Stranford, Sharon A. Kuby's Immunology (2013), W.H. Freeman and Company: New York, 2013

Course Code: HGE.523
Course Title: Techniques in Molecular Medicine
Total Hours: 45

L	T	P	C
3	0	0	3

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Analyze various cellular processes using biochemical techniques.

CLO2: Learn the techniques for the visualization of a broad range of biological processes and features in cell structure.

CLO3: Learn the isolation of nucleic acids and its quantitative & qualitative analysis. Their importance for clinical molecular methods. Apply the knowledge to decipher the mechanisms of molecular and cell biology.

CLO4: Learn various immuno-techniques, Mutation analysis techniques and Cell and tissue culture techniques. Conceptualize principles of these different techniques used in life sciences.

Unit I	11 Hours	CLO1
Microscopy: Light microscopy, phase contrast microscopy, fluorescent microscopy, scanning electron microscopy (SEM/FESEM), transmission electron microscopy (TEM), Atomic force microscopy, Confocal Laser Scanning Microscopy (CLSM), Histochemistry.		

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Unit II Nucleic Acids: Electrophoresis: Principle of gel electrophoresis, polyacrylamide gel electrophoresis (PAGE and SDS-PAGE), agarose gel electrophoresis, pulse field gel electrophoresis (PFGE) and Two-Dimensional gel electrophoresis. Polymerase chain reaction (PCR): Principle, types and applications, PCR based markers: RAPDs, SSRs, SNPs, ISSRs, and SCARs etc. DNA fingerprinting.	11 Hours	CLO2
UNIT III RNA and Proteins: Blotting techniques: Southern, Northern, Western, Dot blotting and hybridization, High throughput techniques: microarray, Next-generation sequencing (NGS): theory and applications, Genome-wide association study (GWAS), Mass Spectrometry, Enzyme Linked Immunosorbent Assay (ELISA).	11 Hours	CLO3
UNIT IV Centrifugation and Chromatography: Basic principles of sedimentation, Types, care and safety aspects of centrifuges, Principles of chromatography, Ion-exchange chromatography, Molecular exclusion (gel filtration) chromatography, Affinity chromatography, Gas-liquid chromatography, High-performance liquid chromatography	12 Hours	CLO4

Transactional Modes: Lecture; Demonstration; Tutorial; Students visit to central instrument facility, Virtual classrooms; Lecture cum demonstration; Problem solving; Self-learning.

Internal assessment shall be through any of the following: Interview on various microscopic techniques; Visit to CIL and compile a report; Case studies solved by DNA fingerprinting, Assignment; In Depth interviews, Analysis of DNA sequencing data, Surprise Tests, term paper, Seminars, discussions and presentations, drawing flow charts for the techniques.

Suggested Readings:

1. Wilson, K. and Walker, J. (2010) Principles and Techniques of Biochemistry and Molecular Biology. Cambridge University Press, Cambridge.
2. Brown, T.A. (2016). Gene cloning and DNA analysis: An Introduction. 6 th Edition, Wiley-Blackwell Publisher, New York.
3. Sue Carson Heather Miller Melissa Srougi D. Scott Witherow (2019). Molecular Biology Techniques. Academic Press, USA
4. Nelson, D. and Cox, M.M. (2016). Lehninger Principles of Biochemistry. W.H. Freeman and Company, New York.
5. Primrose. S.B. and Twyman, R. (2016). Principles of Gene Manipulation and

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Genomics. Blackwell Publishing Professional, U.K.

6. Sambrook, J. (2015). The Condensed Protocols from Molecular Cloning: A Laboratory Manual. Cshl Press. New York.

7. Sambrook, J., Fritish, E.F., & Maniatis, T. (2017). Molecular cloning: A laboratory manual. Cold Spring Harbor Laboratory Press, New York.

Course Code: HGE.524

Course Title: Human Anatomy and Physiology

Total Hours: 45

L	T	P	C
3	0	0	3

Learning outcome

CLO1: Understand anatomical and physiological functions of various tissues.

CLO2: Understand the clinical scenarios and be able to interpret physiological function.

CLO3: Recognize the cell structure and function, histology, gross anatomy and physiology of several organ systems.

CLO4: Understand and predict the body response to stimuli. Recognize the principle of homeostasis and control mechanisms

UNIT I Muscular System: Structure and organization of muscles: skeletal, cardiac and smooth muscles; neuromuscular junction. Cardiovascular System: Physiological anatomy of Heart; cardiac muscle, cardiac cycle; blood constituents; hematopoiesis; cardiovascular regulation.	11 Hours	CLO1
UNIT II Digestive System: Anatomy of Gastrointestinal tract, physiology of salivary secretion, mouth and esophagus, epithelial barrier function; digestion and absorption in GIT; GIT secretions and accessory glands; BMR. Urinary System: Physiological anatomy Kidney; urine formation; regulation of volume and concentration of body fluids, KFT.	12 Hours	CLO2
UNIT III Nervous System: Organization of nervous system, synapse, generation of action potential; vision; hearing and tactile response, degeneration and regeneration of peripheral nerves. Thermoregulation and stress adaptation: Comfort zone; body temperature – physical, chemical and neural regulation; acclimatization.	11 Hours	CLO3
UNIT IV	11 Hours	CLO4

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Respiratory System: Anatomical considerations; mechanism of respiration; neural and chemical regulation of respiration; Physiology of high altitude, hypoxia, PFT.

Reproduction: Physiology of reproductive system (male, female), pregnancy, physiology of fetus.

Transactional Modes: Lecture; Seminar; Tutorial; Virtual classrooms; Problem solving; Self-learning; group discussion.

Internal assessment shall be through any of the following: Assignment; In Depth interviews, Surprise Tests, term paper, Seminars, discussions, and presentations.

Suggested Readings:

1. Brody, T. (1998). Nutritional biochemistry. Academic Press, USA.
2. Devlin, T.M. (2005). Textbook of Biochemistry with clinical correlations. John Wiley & Sons Inc. USA.
3. Hill, R.W, Wyse, G. A. and Anderson, M. (2008). Animal physiology. Sinauer Associates Inc. USA.
4. Khurana. (2006). Textbook of medical physiology. Elsevier India Pvt. Ltd.
5. Murray, R.K. (2009). Harper's illustrated biochemistry. Jaypee Publishers, New Delhi, India.
6. Tyagi, P. (2009). A textbook of Animal Physiology. Dominant Publishers and Distributors, New Delhi, India.
7. Silverthorne D, (2011) Human Physiology, Pearson; 6th edition.
8. Sherman V. (2013) Vander's Human Physiology. McGraw-Hill 13th edition.
9. Jain A.K. (2021). Textbook of physiology. Avichal Publishing Company-9th Edition. Guyton. (2015)., 13th edition Textbook of medical physiology. 11th Edition. Elsevier India Pvt. Ltd. New Delhi.
10. CC Chatterjee's (2022), 14th Edition, Human Physiology, CBS Publishers & Distributors Pvt.Ltd.

Course Code: MME.525

Course Title: Practical Course in Molecular Medicine

Total Hours: 60

L	T	P	C
0	0	8	4

Course Learning Outcomes:

On successful completion of the course the student will be able to:

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CLO1: Gain proficiency in fundamental biotechnological techniques essential for clinical and research applications in biomedical sciences.

CLO2: Acquire expertise in advanced molecular biology techniques for quantitative gene expression, protein identification and quantification.

CLO3: Master histological techniques and immunohistochemical analysis, coupled with the microscopic identification of lymphoid organs, to achieve comprehensive skills in tissue preparation, staining, and the analysis of cellular and tissue morphology.

CLO4: Develop proficiency in cell culturing and viability assessment, enabling the evaluation of cell proliferation, cytotoxicity, and metabolic activity for research and therapeutic applications.

1.	Separation of Plasma and Serum from the blood samples.	CLO1
2.	Blood film preparation and identification of cells	
3.	Polymerase Chain Reaction	
4.	Agarose Gel Electrophoresis	CLO2
5.	Real-Time PCR	
6.	Gel preparation and transfer	
7.	Tissue fixation and sectioning	CLO3
8.	H and E staining of tissues	
9.	Microscopic identification of Lymphoid organs	CLO4
10.	Cell Culturing	

Transactional Modes: Lecture; Demonstration; Virtual classrooms; Problem solving; Self-learning; YouTube demonstrations; Lab performances.

Evaluation criteria for practical courses:

- Continuous Assessment = **60 Marks**

Based on performance and good lab practices of the students and completion of the record book (60 divided by the number of practical's)

- Final Examination = **40 Marks**

Subjective question = 10 Marks

Performing experiment = 20 Marks

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Viva voce = 10 Marks

Interdisciplinary Courses

Course Code: HGE.529
Course Title: Introduction to Intellectual Property Rights
Total Hours: 30

L	T	P	C
2	0	0	2

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Understand the concept and genesis of IP

CLO2: Differentiate between plagiarism and fair use of copyright material

CLO3: Understand the Patent regime of India

CLO4: Conceptualize the other forms of IP namely Trademark, Industrial Design, Trade Secret, New varieties of plant

UNIT I Hours Brief history, current status and career opportunities in IP Introduction to IP, Genesis and development of concept of IPR; WIPO administered Treaties: Paris Convention, 1883, the Berne Convention, 1886, the TRIPS Agreement, 1994; the WIPO Convention, 1967; National Innovation and Startup Policy for Students and Faculty 2019, Career Opportunities in IP.	8	CLO1
UNIT II Hours Copyright and Layout Design Protection Copyright and related rights; Plagiarism; Fair Use of copyright material; Layout Design Protection.	6	CLO2
UNIT III Hours Patent Regime in India: Patents, patentability of inventions; non-patentable subject matter, Patent registration procedure in India; Protection of Traditional Knowledge, Assignment and license of patented technology; Patent filing routes for other countries: Convention Application and Patent Cooperation Treaty (PCT) application.	8	CLO3
UNIT IV Hours Other forms of IP	8	CLO4

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Concept, Registration and term of protection: Trademark, Industrial Design, Trade Secret, Protection of New varieties of plant, Geographical Indications	
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Transactional Modes: Lecture; Demonstration; Tutorial; Virtual classrooms; Lecture cum demonstration; Problem solving; Self-learning, Class activity based

Internal assessment shall be through any of the following: Surprise Tests, one sentence summary, case analysis, simulated problem solving, open book techniques, classroom assignments, term paper, Seminars.

Suggested Readings:

1. Dutfield G. (2003). Intellectual Property Rights and the Life Science Industries: A Twentieth Century History (Globalization and Law). Routledge.
2. Mahop, M.T. (2010). Intellectual Property, Community Rights and Human Rights: The Biological and Genetic Resources of Developing Countries. Routledge.
3. Khor M. (2002). Intellectual Property, Biodiversity and Sustainable Development: Resolving the Difficult Issues. Zed Books limited.
4. Ahuja, V.K. (2017). Law relating to Intellectual Property Rights. LexisNexis, India. 3rd Edition.
5. Mahop, M.T. (2010). Intellectual Property, Community Rights and Human Rights: The Biological and Genetic Resources of Developing Countries. Routledge, USA.
6. Neeraj, P. and Khusdeep, D. (2014). Intellectual Property Rights. India, IN: PHI learning Private Limited.
7. Nithyananda, K V. (2019). Intellectual Property Rights: Protection and Management. India, IN: Cengage Learning India Private Limited.

Weblinks:

1. World Intellectual Property Organization (<https://www.wipo.int/about-ip/en/>)
2. Office of the Controller General of Patents, Designs & Trademarks (<http://www.ipindia.nic.in/>)

Discipline Elective-II

Course Code: MME.527

Course Title: Molecular Endocrinology and Signal Transduction

Total Hours: 45

L	T	P	C
3	0	0	3

Course Learning Outcomes:

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On successful completion of the course the student will be able to:

CLO1: Know endocrine system and signal transduction at physiological levels

CLO2: Conceptualize and understand the endocrine cellular signaling

CLO2: Understand various human hormones.

CLO3: Develop understanding regarding disorders of the immune system.

Unit: 1 Hours Endocrine glands, and hormones as chemical messengers Regulation of hormone production and release: change in homeostasis, sensory stimulus and others.	10	CLO1
Unit: 2 Hours Hormones: Structures, Receptor type, Regulation of biosynthesis and release (including feedback mechanism), Physiological and Biochemical actions, Pathophysiology (hyper & hypo secretion). Hypothalamic Hormones: CRH, TRH, GnRH, PRL/PRIH, GHRH/GHRIH. Pituitary Hormones - Anterior Pituitary hormones- Growth hormone, Prolactin, POMC peptide family, LH, FSH, TSH; Posterior Pituitary: Vasopressin, Oxytocin, reproductive hormones, Other organs with endocrine function: Heart (ANP), Kidney (erythropoietin), Liver (Angiotensinogen, IGF-1), Adipose tissue (Leptin, adiponectin).	15	CLO2
Unit: 3 Hours Cell Signaling and Mechanism of Hormone Action: G protein linked receptor family; Signal transduction pathways involving G-proteins, Adenylcyclases, Ca ²⁺ , Phosphoinositides, PI-3 Kinase, DAG, cAMP, cGMP, NO, Protein kinases (A,B,C,G), Phosphoprotein phosphatases & Phosphodiesterases. Receptor tyrosine kinase family- EGF receptor family, Insulin receptor family, & Cytokine/erythropoietin receptor family associated with non-receptor Tyrosine kinase (Signal transduction pathways involving: SH2 proteins, Ras, IRS-1, Raf, MEK, MAP kinase, JAK-STAT pathway).	10	CLO3
Unit: 4 Hours Endocrine disorders: Gigantism, Acromegaly, dwarfs, pigmies; Pathophysiology: Diabetes insipidus. Thyroid Hormone (include biosynthesis) Goiter, Graves' disease, Cretinism, Myxedema, Hashimoto's disease. Pancreatic Hormones: Insulin, Glucagon, Diabetes type I & II. Hormones associated with obesity: Ghrelin, Leptin.	10	CLO4

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Transactional Modes: Lecture; Demonstration; Tutorial; Lecture cum demonstration; Problem solving; Self-learning.

Internal assessment shall be through any of the following: Surprise Tests, case analysis, simulated problem solving, classroom assignments, term paper, presentations and discussions.

Suggested Reading:

1. Norris, D.O., & Carr, J.A. (2017). Vertebrate Endocrinology. Academic Press.
2. Widmaier, E.P., Raff, H., & Strang, K.T. (2013). Vander's Human Physiology. McGraw-Hill Higher Education.
3. Ari Sitaramayya (2012) Introduction to Cellular Signal Transduction (Hormones in Health and Disease), Springer science, New York.
4. Bastien D. G., IJsbrand M. K. and Peter E.R. T. (2015), 3rd edition, Signal Transduction, Academic Press, MA, USA

Course Code: HGE.527
Course Title: Human Embryology and Developmental Genetics
Total Hours: 45

L	T	P	C
3	0	0	3

Course Learning Outcomes:

On successful completion of the course the student will be able to:

- CLO1: Conceptualize basics of reproductive physiology
 CLO2: Correlate genetic regulation in different embryonic developmental stages
 CLO3: Evaluate the role of biomolecules in embryonic development.
 CLO4: Know different genetic and environmental triggers for post-natal development, ageing and senescence.

Unit: I Gametes, fertilization and early development: Gametogenesis; fertilization; types of cleavage; morphogenesis; gastrulation: cell movement and formation of germ layers in frog, chick and mouse; fate map; fetal membrane and placenta; potency, commitment, specification, induction, competence, determination, and differentiation.	12 Hours	CLO1
Unit: II Gene expression regulation in development: Basics of gene expression regulation during early embryogenesis; homeotic genes, P granules, role of key developmental genes: Polycomb repressor complexes (PRC1 and PRC2), Sox proteins.	11 Hours	CLO2
Unit: III	11 Hours	CLO3

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Stem Cell and Organogenesis: Stem cell: embryonic and adult; cell-cell communication; neural crest cells and axonal specificity; vertebrate eye and central nervous system development; hematopoiesis.		
Unit: IV	11 Hours	CLO4
Genetics of pattern and organ formation: Chromatin diminution; endoreplication cycles; gene amplification; Drosophila: formation of body axis, segmentation genes, homeotic gene function; Vertebrates: axes formation and HOX genes.		

Transactional Modes: Lecture; Demonstration; Tutorial; Virtual classrooms; Lecture cum demonstration; Problem solving; Self-learning.

Internal assessment shall be through any of the following: Surprise Tests, one sentence summary, case analysis, simulated problem solving, open book techniques, classroom assignments, homework assignments, term paper, Seminars, term papers, discussions, and presentations.

Suggested Readings:

1. Gilbert, S.F. (2013). Developmental Biology. Tenth Edition.
2. Slack, J.M.W. (2012). Essential Developmental Biology. Third Edition.
3. Moody, A.A. (2014). Principles of Developmental Genetics. Second Edition.
4. Slack, J.M.W. (2018). The Science of Stem Cells. First Edition.
5. Milunsky, J. & Milunsky, A. (2010). Genetic Disorders and the Fetus: Diagnosis, Prevention & Treatment. Willey Blackwell India, New Delhi.
6. Prakash, G. (2007). Reproductive Biology. Narosa Publication House Pvt. Ltd., New Delhi.
7. Sadler, T.W., Tosney, K., Chescheir, N.,C., Imseis, H., Leland, J. and Sadler-Redmond, S.,L. (2011). Langman's Medical Embryology (Longmans Medical Embryology). Lippincott Williams and Wilkins.
8. Keith L. Moore BA, T. V. N. Persaud MD., Mark G. Torchia (2019) The Developing Human Clinically Oriented Embryology, Elsevier, Netherlands.

Semester-III

Course Code: MME.551

Course Title: Molecular Basis of Human Diseases

Total Hours: 45

L	T	P	C
3	0	0	3

Course Learning Outcomes:

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On successful completion of the course the student will be able to:

CLO1: Acquire knowledge on basic mechanisms of non-communicable human diseases.

CLO2: Understand the molecular basis of human genetic disorders.

CLO3: Gain knowledge about molecular mechanisms underlying the pathogenesis of communicable human diseases.

CLO4: Know modern therapeutic approaches in development/clinical practice.

<p>Unit: 1 Hours</p> <p>Non-Communicable Diseases: Molecular basis of: Diabetes, Chronic obstructive pulmonary disease, Cardiomyopathies, Hypertension, Cancer and neuronal disorders such as Autism, Alzheimer's and Parkinson. Schizophrenia, Mental Retardation, Major depressive disorder, Anxiety disorder</p>	11	CLO1
<p>Unit: 2 Hours</p> <p>Genetic disorders: Classifications of genetic disorders, Intersex Disorders: Male Pseudo-hermaphrodite (MPH), Female Pseudo-hermaphrodite (FPH), True Hermaphrodites (TH), Mixed gonadal dysgenesis (MGD) & Dysgenetic male pseudohermaphrodite (DMP) and Persistent Mullerian duct syndrome (PMDS), Sickle cell anemia, Thalassemia's and Hemophilia's and Hematopoietic Malignancies. Muscular Dystrophy. Glycogen Storage Diseases (Pompe disease, Tay Sach disease, Niemann-Pick disease). Cystic Fibrosis, Down Syndrome, Huntington's Disease, Colour Blindness.</p>	12	CLO2
<p>Unit: 3 Hours</p> <p>Communicable Diseases: Mechanisms of Infection and Therapeutic Interventions of: HIV/AIDS, SARS, TB and Hepatitis. Molecular basis of antimicrobial resistance and its detection. Molecular approaches in clinical microbiology, antimicrobial agents; Sulfa drugs; Antibiotics: Penicillin and Cephalosporins; Broad-spectrum antibiotics; Antibiotics from prokaryotes; Antifungal antibiotics.</p>	12	CLO3
<p>Unit: 4 Hours</p> <p>Novel therapies for diseases: Tyrosine kinase inhibitor, Monoclonal antibody, Chemo & Radio, Gene Therapies, Small peptides. Limitations, ethical and biosafety issues in gene therapies.</p>	10	CLO4

Transactional Modes: Lecture; Demonstration; Tutorial; Lecture cum demonstration; Problem solving; Self-learning.

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Internal assessment shall be through any of the following: Surprise Tests, case analysis, simulated problem solving, classroom assignments, term paper, presentations, and discussions.

Suggested Reading:

1. Patch, H. S. C. (2017). Genetics for the Health Sciences. Scion Publishing Ltd., UK.
2. Brown, S. M., (2018). Essentials of Medical Genomics. Wiley-Blackwell.
3. Jocelyn, E. K., Elliot, S. G., & Stephen, T. K. (2018), Lewin's Gene X. Jones & Barlett Publishers.
4. Milunsky, A., & Milunsky, J. (2015). Genetic Disorders and the Fetus: Diagnosis, Prevention and Treatment, 6th Edition. Wiley-Blackwell publishers.
5. Trent, R. J. (2017). Molecular Medicine: Genomics to Personalized Healthcare. Academic Press.
6. Trent, R. J. (2015). Molecular Medicine: An Introductory Text. Academic Press.
7. Elles, R., & Mountford, R. (2012). Molecular Diagnosis of Genetic Diseases Series: Methods in Molecular Medicine.
8. Coleman, W. B., & Tsongalis, G. J. (2019). The Molecular Basis of Human Disease. Academic Press.
9. Nussbaum, R.L., McInnes, R. Mc., & Willard, H.F. (2017). Genetics in Medicine. Elsevier Inc., Philadelphia.
10. Read, A., & Donnai D. (2017). New clinical Genetics. Scion Publishing Lmt., Oxfordshire, UK.
11. Kandel, E. R., Schwartz, J. H., & Jessell, T. M. (2021), 6th edition, Principles of neural science. McGraw-Hill, Health Professions Division.

Course Code: MME.552

Course Title: Genetic Engineering and Recombinant Therapeutics

Total Hours: 45

L	T	P	C
3	0	0	3

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Know about genetic engineering and its applications.

CLO2: Gain knowledge about various cloning, expression vectors and their importance in research.

CLO3: Understand concept of artificial chromosomes and their potential applications.

CLO4: Learn therapeutics aspect of recombinant DNA technologies

Syllabus

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Unit: 1 Hours Basics of Genetic Engineering: Gene manipulation tools for molecular cloning, restriction enzymes their types, cohesive and blunt and ligation, linkers, adaptors, homopolymeric tailing, transformation, transfection: chemical and physical methods, sequencing methods, gene cloning, in-silico methods of design.	11	CLO1
Unit: 2 Hours Gene Cloning Vectors: Plasmids, bacteriophages, cloning in M13 Vectors, phagemids, Lambda vectors; insertion and replacement vectors, Cosmid vectors, Artificial chromosome vectors (YACs, BACs), Animal virus derived vectors-Sv-40, retroviral vectors, Expression vectors	11	CLO2
Unit: 3 Hours Techniques in Genetic Engineering: Isolation and Detection of DNA, RNA and proteins by Southern blotting, Northern blotting, Western blotting and in situ hybridization, Site Directed Mutagenesis, Yeast two hybrid system, phage display, characterization of expressed proteins through various biophysical, biochemical methods,	11	CLO3
Unit: 4 Hours Applications of recombinant DNA technology: Applications of rDNA in diagnosis of pathogens and abnormal genes, Gene Editing through CRISPR/CAS system, transgenic animals for production of proteins and pharmaceuticals, Biosafety and Ethical considerations in genetic engineering.	12	CLO4

Transactional Modes: Lecture; Demonstration; Tutorial; Lecture cum demonstration; Problem solving; Self-learning.

Internal assessment shall be through any of the following: Surprise Tests, case analysis, simulated problem solving, classroom assignments, term paper, presentations, and discussions.

Suggested Reading:

1. R.W. Old., & S.B. Primrose. (2017). Principles of Gene Manipulation Blackwell science.
2. Bernard R. Glick., & Jack J. Pasternak. (2018) Molecular Biotechnology ASM Press Washington.
3. James, Watson Micheal Gilman Jan Witkowsk (2017) Recombinant DNA, CSHL, New York.

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4. Cokin, R., & Bjorn, C. (2016). Basic Biotechnology Cambridge University press.
5. John E. Smith. (2019). Biotechnology by Cambridge University press.
6. Watson, J. D., Baker, T. A., Bell, S. P., Gann, A., Levine, M., & Losicke, R. (2017). Molecular Biology of Gene by Watson CSHL Press New York.
7. Sambrook, J & Sambrook, R. (2018). Molecular cloning, CSHL Press, New York.

Course Code: MME.553

Course Title: Trends in Molecular Medicine

Total Hours: 30

L	T	P	C
3	0	0	3

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Know about background of Molecular Medicine.

CLO2: Know about development of therapeutics.

CLO3: Understand cellular microenvironment and problems of drug resistance.

CLO4: Develop knowledge about translational research.

Unit: 1 Hours Introduction to Molecular Medicine, contribution of genomics, transcriptomics and proteomics in human diseases, developing novel biomarkers and therapies using high throughput technologies.	10	CLO1
Unit: 2 Hours Molecular Medicine Therapeutics: Gene therapy and recombinant molecules in medicine and therapeutic development, pharmacogenomics.	10	CLO2
Unit: 3 Hours Signal Transduction and its Role in Human Diseases: cell signaling and human diseases, Cellular and tissue microenvironment in diseases, drug resistance with convention chemotherapies	14	CLO3
Unit: 4 Hours Advances in translational research: nano-biotechnology and its applications in molecular medicine, immunotherapies in human diseases, translational research and its contributions in disease therapeutics.	11	CLO4

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Transactional modes: Lecture; Demonstration; Tutorial; Lecture cum demonstration; Problem solving; Self-learning.

Internal assessment shall be through any of the following: Surprise Tests, one sentence summary, case analysis, simulated problem solving, open book techniques, classroom assignments, homework assignments, term paper, presentations and discussions.

Suggested Reading:

1. Littwack, G. (2018). Human Biochemistry and Disease. Academic Press.
2. Trent, R. J. (2016). Molecular Medicine, Fourth Edition: Genomics to Personalized Healthcare. Academic Press.
3. Trent, R. J. (2015). Molecular Medicine: An Introductory Text. Academic Press.
4. Elles, R., Mounfield, R. (2011). Molecular Diagnosis of Genetic Diseases. Springer Publication.
5. Liciniio, J., & Wong, M. L. (2018). Pharmacogenomics: The Search for Individualized Therapies. Wiley.

Course Code: HGE.554

Course Title: Tools of Bioinformatics (Practical)

Total Hours: 60

L	T	P	C
0	0	6	3

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Analyze publicly available and experimental genomics data

CLO2: Analyze sequence alignment and generation of PCR oligos

CLO2: Perform in-silico prediction of protein structures and interactions

CLO3: Perform next generation sequence and microarray data analysis and functional annotation of genetic findings.

CLO4: Demonstrate physical property of biomolecules in silico

1.	Access to sequence databases (downloading DNA/RNA/Protein sequences).	CLO1
2.	Genomics/transcriptomics/proteomics data retrieval and analysis	
3.	Primer designing for PCR and RT-PCR	CLO2
4.	Performing sequence alignment using various tools (BLAST, MSA)	
5.	Pipeline of Next generation RNA-seq/DNA sequencing analysis.	CLO3
6.	Genome wide association study and DNA microarray-data analysis	

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7.	Analysis of linkage disequilibrium plots.	
8.	Prediction of Protein structure using sequence database	CLO4
9.	Practical insights of tertiary structure prediction and comparative modeling	
10.	Protein-protein and protein-ligand docking	

Transactional Modes: Hands-on training; Demonstration; Tutorial; Lecture cum demonstration; Problem solving; Self-learning.

Internal assessment shall be through any of the following: Surprise Tests, one sentence summary, case analysis, simulated problem solving, open book techniques, classroom assignments, homework assignments, term paper, Seminars, term papers, discussions, and presentations.

Evaluation criteria for practical courses:

- Continuous Assessment = **60 Marks**

Based on performance and good lab practices of the students and completion of the record book (60 divided by the number of practicals)

- Final Examination = **40 Marks**

Subjective question = 10 Marks

Performing experiment = 20 Marks

Viva voce = 10 Marks

Suggested Readings:

1. Laboratory Protocols. Third Edition. 2005. CIMMYT Applied Molecular Genetics Laboratory.
2. Koliantz, G & Szymanski, D.B. Genetics A Laboratory Manual. Second Edition. 2009. American Society of Agronomy, Crop Science Society of America.
3. Sambrook, J & Green, M.R. Molecular Cloning: A laboratory manual. Fourth Edition. 2013. Cold Spring Harbor Laboratory Press, U.S.
4. Pazos, F & Chagoyen, M. Practical protein bioinformatics.2015. Springer International Publishing, Switzerland.
5. Agostino, M. Practical Bioinformatics. 2012. Garland Science. Taylor & Francis Group. New York and London.

Discipline Elective-III

Course Code: MME.555

Course Title: Evolution and Developmental Biology

L	T	P	C
3	0	0	3

Syllabus

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Total Hours: 45

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Gain knowledge on concept related to origin of life.

CLO2: Develop understanding regarding molecular evolution and original of first cell.

CLO3: Know about concepts related to developmental processes.

CLO4: Understand pathology related to mechanisms of development and differentiation.

Unit: 1 Hours Origin of Life: History of evolutionary ideas, Modern evolution theory, Natural Selection, Adaptation, The origin of species. <i>Nature of Evolution (Microevolution & macroevolution), Punctuated equilibria & Phyletic gradualism and Pattern of Evolution (Anagenesis & Cladogenesis).</i>	11	CLO1
Unit: 2 Hours Paleontology and Molecular Evolution: The evolutionary time scale, Major events in the evolutionary time scale, Stages in primate evolution, Human evolution, Neutral evolution, Molecular divergence and molecular clocks, Gene duplication and divergence.	11	CLO2
Unit: 3 Hours Basic Concepts of Development: Potency, Commitment, Specification, Induction, Competence, Determination and Differentiation, Morphogenetic gradients, Cell fate and cell lineages, Stem cells, Genomic equivalence and the cytoplasmic determinants, Imprinting, Mutants and transgenics in analysis of development.	12	CLO3
Unit: 4 Hours Gametogenesis, Fertilization, embryology and neurulation: Production of gametes, Cell surface molecules in sperm-egg recognition; embryonic development and formation of germ layers in humans, fetal development, sex determination, neural tube formation.	11	CLO4

Transactional Modes: Lecture; Demonstration; Tutorial; Lecture cum demonstration; Problem solving; Self-learning.

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Internal assessment shall be through any of the following: Surprise Tests, one sentence summary, case analysis, simulated problem solving, open book techniques, classroom assignments, homework assignments, term paper, Seminars, term papers, discussions, and presentations.

Suggested Reading:

1. Darwin, C. (1956). The Origin of Species: By means of natural selection or the preservation of favored races in the struggle for life (No. 575.0162). Oxford University Press.
 2. Dawkins, R. (1996). The blind watchmaker: Why the evidence of evolution reveals a universe without design. WW Norton & Company.
 3. Futuyma, D.J. (2017). Evolution. Sinauer Associates Inc. USA.
 4. Wilt, F. H., & Hake, S. (2004). Principles of developmental biology. W.W. Norton & Company, New York, USA.
 5. Hall, B.K., & Hallgrimsson, B. (2017). Strasburger's Evolution. Jones and Bartlett Publishers, India.
 6. Lewin, R. (2014). Human Evolution - An Illustrated Introduction. Wiley-Blackwell, USA.
 7. Scott, F., & Gilbert, S.F. (2017). Developmental Biology. Sinauer Associates, Inc. USA.
 8. Slack, J.M.W. (2015). Essential Developmental Biology, Wiley-Blackwell, USA.
- Barton N. H. (2007), Evolution, Cold Spring Harbor Laboratory Press, USA
Stillman Bruce (2009), Evolution: The molecular landscape, Cold Spring Harbor Laboratory Press, USA

Course Code: HGE.555

Course Title: Biosafety, Bioethics, and Intellectual Property Rights

Total Hours: 45

L	T	P	C
3	0	0	3

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Interpret the bioethical issues concerning biotechnological advancements like recombinant DNA technology, cloning, gene manipulation.

CLO2: Implement biosafety while carrying out research.

CLO3: Distinguish different types of Intellectual Property Rights.

CLO4: Describe the ways of protecting traditional knowledge from Bio piracy.

UNIT I	11 Hours	CLO1
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Biosafety: Good laboratory practices; Risk and safety assessment from genetically engineered organisms; special procedures for r-DNA based products; biological containment (BC) and physical containment (PC); CDC biosafety levels; biohazard management.		
UNIT II Bioethics: Ethical considerations during research, Use of Animals for clinical research, Embryonic and adult stem cell research, assisted reproductive technologies, cloning, MTP and Euthanasia; the element of informed consent; ethical issues of the human genome project.	11 Hours	CLO2
UNIT III Intellectual Property Rights (IPRs): Various forms of IP – patents, industrial designs, trademark, geographical indications, and plant breeder's right; copyright: fair use, plagiarism; protection of indigenous intellectual property.	11 Hours	CLO3
UNIT IV Patent system: Patent filing procedure in India and ways of patent protection in other countries: Determination of patentability of inventions, filing a patent application in India: timeline, procedure involved in the granting of a patent, patent cooperation Treaty (PCT).	12 Hours	CLO4

Transactional Modes: Lecture; Demonstration; Virtual classrooms; Tutorial; Lecture cum demonstration; Problem solving; Case study; Self-learning.

Internal assessment shall be through any of the following:

Surprise Tests, one sentence summary, case analysis, simulated problem solving, open book techniques, classroom assignments, homework assignments, term paper, Seminars, term papers, discussions, and presentations.

Suggested Readings:

1. Clarke, A (2012). Genetic Counseling: Practice and Principles. Taylor & Francis
2. Fleming, D.O. and Hunt, D.L. (2006). Biological Safety: Principles and Practices. American Society for Microbiology, USA.
3. Mahop, M.T. (2010). Intellectual Property, Community Rights and Human Rights: The Biological and Genetic Resources of Developing Countries. Routledge.
4. Shannon, T.A. (2009). An Introduction to Bioethics. Paulist Press, USA.
5. Thompson J and Schaefer, B.D (2013). Medical Genetics: An Integrated Approach. McGraw Hill.
6. Vaughn, L. (2009). Bioethics: Principles, Issues, and Cases. Oxford University Press, UK.
7. WHO. (2005). Laboratory Biosafety Manual. World Health Organization.
8. Ahuja, V.K. (2017). Law relating to Intellectual Property Rights. LexisNexis, India. 3rd Edition.

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9. Mahop, M.T. (2010). Intellectual Property, Community Rights and Human Rights: The Biological and Genetic Resources of Developing Countries. Routledge, USA.
10. Neeraj, P. and Khusdeep, D. (2014). Intellectual Property Rights. India, IN: PHI learning Private Limited.
11. Nithyananda, K V. (2019). Intellectual Property Rights: Protection and Management. India, IN: Cengage Learning India Private Limited.

Value added Course

Course Code: HGE.529
Course Title: Principles of Ecological Science
Total Hours: 30

L	T	P	C
2	0	0	2

Course Learning Outcomes:

On successful completion of the course the student will be able to:

CLO1: Improve their knowledgebase about the basics of ecological components

CLO2: Conceptualize the ecosystem and its components

CLO3: Contribute to for population ecology and conservation biology

CLO4: Improve student's aptitude for ecological science research and development

UNIT I Environment components: Physical environment (Climate, atmosphere, light, soil, temperature); biotic environment; biotic and abiotic interactions. Ecological amplitude; Ecological adaptation; Ecological species concept; Ecotype & Ecads; K-selected & r-selected species; Concept of habitat and niche; character displacement.	6 Hours	CLO1
UNIT II Ecosystem and Ecological Succession: Ecosystem components; Productivity & energy flow; Food chain; Food web; Concepts of the trophic level, ecological pyramid and mineral cycling (C, N, P); primary production and decomposition; Kinds of ecosystems: terrestrial and aquatic; Biomes. Ecological Successions: Types; mechanisms; changes involved in succession; concept of climax.	8 Hours	CLO2
UNIT III Population Ecology and Conservation Biology: Characteristics of a population; Population dispersal; Population Growth; Biotic community: Ecological characteristics; Nature & Structure of community; Ecological	8 Hours	CLO3

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interdependence & interaction; Biodiversity: Levels of diversity and its measurement; Components & gradients of biodiversity; Conservation of biodiversity; Project Tiger; Biogeographic classification of India.		
UNIT IV Applied Ecology: Environmental pollution; Air pollution: Pollutants, greenhouse effect, global warming, Kyoto and Montreal Protocol, ozone, acid rain. Water pollution: Biochemical oxygen demand, chemical oxygen demand, eutrophication; Soil pollution; Bioremediation, phytoremediation.	8 Hours	CLO4

Transactional Modes: Lecture; Demonstration; Tutorial; Lecture cum demonstration; Problem solving; Self-learning; MCQ practice.

Internal assessment shall be through any of the following: Surprise Tests, one sentence summary, case analysis, simulated problem solving, open book techniques, classroom assignments, homework assignments, term paper, Seminars, term papers, discussions, and presentations.

Suggested Readings:

1. Smith, Robert Leo, et al. "Elements of ecology." (2015): E9.
2. Recknagel F. 2002 Ecological Informatics: Understanding Ecology by Biologically- Inspired Computation, Springer, New York.
3. Odum E.P. 1983 Basic Ecology. Saunders International Edition, Japan
4. Michael Begon, 2020 Ecology: From Individuals to Ecosystems 5th Edition,

Foundation Course

Course Code: HGE.558

Course Title: Innovation and Entrepreneurship

Total Hours: 15

L	T	P	C
2	0	0	2

Course Learning Outcomes:

On the completion of this course, the learners will:

CLO1: Understand the basic concepts of entrepreneur, entrepreneurship, and its importance.

CLO2: Aware of the issues, challenges, and opportunities in entrepreneurship.

CLO3: Develop capabilities of preparing proposals for starting small businesses.

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CLO4: Know the availability of various institutional supports for making a new start-up.

Unit I Nature, characteristics, functions and its role in economic development; entrepreneurial behavior and skills; the role of women entrepreneurs and social entrepreneurs in society; market driven and technology driven entrepreneurship, startups	4 Hours	CLO1
Unit II Business plan and funding opportunities Business model canvas; supply and customer chains, Business to business (B2B), Direct to customer (D2C) business types and funding opportunities: marketing and sales strategy; public funded schemes and angel funding	4 Hours	CLO2
Unit III Blending University Research and Entrepreneurship culture: Pre Incubation and Incubation facilities, Idea hackathons, Policy to encourage entrepreneurship in students and faculty; National Innovation and startup policy for students and faculty 2019 . Case studies	4 Hours	CLO3
Unit IV Intellectual Property (IP) and Technology Transfer: Different forms of Industrial intellectual property (IP)-patent, industrial design and trademark, technology transfer requirements and process	4 Hours	CLO4

Suggested Readings:

1. Arora, Renu (2008). Entrepreneurship and Small Business, Dhanpat Rai & Sons Publications.
2. Chandra, Prasaaan (2018). Project Preparation, Appraisal, Implementation, Tata Mc-Graw Hills.
3. Desai, Vasant (2019). Management of a Small-Scale Industry, Himalaya Publishing House.
4. Jain, P. C. (2015). Handbook of New Entrepreneurs, Oxford University Press.
5. Srivastava, S. B. (2009). A Practical Guide to Industrial Entrepreneurs, Sultan Chand & Sons.
6. National Innovation and startup policy for students and faculty 2019, Government Policy document <http://rmkcet.ac.in/RMK/NISP%20policy.pdf>

Course Code: MME.600

L	T	P	C
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Syllabus

MSc in Molecular Medicine (Batch: 2024-2026)

Course Title: Dissertation Part-I

0	0	8	4
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Total Hours: 120

Course Learning Outcomes:

On successful completion of the course the student will be able to:

1. Apply genetic and genomics technique for research.
2. Construct study design
3. Interpret result of a genetic experiment.
4. Present oral and written scientific communication skills.

The objective of dissertation part-I would be to ensure that the student learns the nuances of the scientific writing. Herein the student shall have to write his synopsis including an extensive review of literature with simultaneous identification of scientifically sound (and achievable) objectives backed by a comprehensive and detailed methodology. Student may start working in the respective laboratory.

Students can opt for dissertation work in industry, national institutes or Universities in the top 100 NIRF ranking. Group dissertation can be opted, with a group consisting of a maximum of four students. These students may work using a single approach or multidisciplinary approach. Research projects can be taken up in collaboration with industry or in a group from within the discipline or across the discipline

Synopsis will be evaluated as per the University policy.

Evaluation criteria:

Examiner	Marks	Evaluation
Supervisor	50	Dissertation proposal and presentation
HoD and Senior-most faculty of the department	50	Dissertation proposal and presentation

Syllabus

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Semester IV

Course Code: MME.601
Course Title: Dissertation Part-II
Total Hours: 600

L	T	P	C
0	0	40	20

Course Learning Outcomes:

On successful completion of the course the student will be able to:

1. Apply genetic and genomics technique for research
2. Design research studies and perform research following scientific and ethical guidelines
3. Interpret results of genetic and molecular experiments
4. Present oral and written scientific literatures

During the course student will perform:

- a. Synthesis of research hypothesis
- b. Review of literature and identify research gap
- c. Formulate methodology to achieve the objective of the research idea
- d. Present articles and research ideas to fellow students and in other platforms
- e. Perform research and interpret the results
- f. Write research reports and may publish research findings (if significant)

Dissertation will be evaluated by the Department, as per the University policy.
Evaluation criteria

Examiner	Marks	Evaluation
Supervisor	50	Continuous assessment (regularity in work, mid-term evaluation) dissertation report, presentation, final viva-voce
External expert, HoD and senior-most faculty of the department	50	Dissertation report (30), presentation (10), final viva-voce (10)

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Examination pattern

Core, Discipline Elective, Compulsory Foundation, Value Added and Interdisciplinary Courses			Discipline Enrichment Course		Entrepreneurship Course	
	Marks	Evaluation	Marks	Evaluation	Marks	Evaluation
Internal Assessment	25	Various	-	-	-	-
Mid-semester test (MST)	25	Subjective	50	Objective	25	Objective
End-semester test (EST)	50	Subjective (70%) Objective (30%)	50	Objective	25	Subjective

Dissertation Proposal (Third Semester)			Dissertation (Fourth Semester)		
	Marks	Evaluation		Marks	Evaluation
Supervisor	50	Dissertation proposal and presentation	Supervisor	50	Continuous assessment (regularity in work, mid-term evaluation) dissertation report, presentation, final viva-voce
HoD and senior-most faculty of the department	50	Dissertation proposal and presentation	External expert, HoD and senior-most faculty of the department	50	Dissertation report (30), presentation (10), final viva-voce (10)

Evaluation pattern similar to fourth semester dissertation will apply for internship where supervisor will award 50% marks and external co-supervisor, HoD and senior-most faculty will award 50% marks.

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